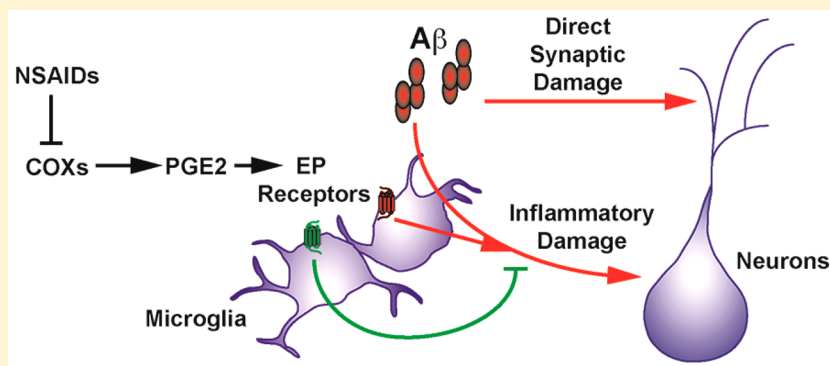


# Untangling the Web: Toxic and Protective Effects of Neuroinflammation and PGE<sub>2</sub> Signaling in Alzheimer's Disease

Nathaniel S. Woodling<sup>†</sup> and Katrin I. Andreasson\*

Department of Neurology and Neurological Sciences, Stanford University School of Medicine, 1201 Welch Road, Stanford, California 94305, United States



**ABSTRACT:** The neuroinflammatory response has received increasing attention as a key factor in the pathogenesis of Alzheimer's disease (AD). Microglia, the innate immune cells and resident phagocytes of the brain, respond to accumulating A $\beta$  peptides by generating a nonresolving inflammatory response. While this response can clear A $\beta$  peptides from the nervous system in some settings, its failure to do so in AD accelerates synaptic injury, neuronal loss, and cognitive decline. The complex molecular components of this response are beginning to be unraveled, with identification of both damaging and protective roles for individual components of the neuroinflammatory response. Even within one molecular pathway, contrasting effects are often present. As one example, recent studies of the inflammatory cyclooxygenase–prostaglandin pathway have revealed both beneficial and detrimental effects dependent on the disease context, cell type, and downstream signaling pathway. Nonsteroidal anti-inflammatory drugs (NSAIDs), which inhibit cyclooxygenases, are associated with reduced AD risk when taken by cognitively normal populations, but additional clinical and mouse model studies have added complexities and caveats to this finding. Downstream of cyclooxygenase activity, prostaglandin E<sub>2</sub> signaling exerts both damaging pro-inflammatory and protective anti-inflammatory effects through actions of specific E-prostanoid G-protein coupled receptors on specific cell types. These complexities underscore the need for careful study of individual components of the neuroinflammatory response to better understand their contribution to AD pathogenesis and progression.

**KEYWORDS:** Alzheimer's disease, microglia, amyloid  $\beta$ , neuroinflammation, cyclooxygenases, prostaglandin E<sub>2</sub>, EP2 receptor, EP3 receptor, and EP4 receptor

Alzheimer's disease (AD) is the most common cause of dementia among the elderly, with a prevalence of one in eight people over the age of 65. With projected demographic shifts toward an aging population, the annual incidence of new AD cases is projected to triple by 2050,<sup>1</sup> with costs of care rising from \$226 billion in 2015 to more than \$1 trillion by 2050 (Alzheimer's Association Facts, 2015). This coming epidemic represents one of the most serious challenges to our health care system in the coming decades. There is an urgent need for strategies to treat and, perhaps even more importantly, to prevent AD. Given that AD prevalence doubles every 5 years after the age of 65, strategies delaying the onset of AD by even 5 years would be predicted to reduce disease burden by 50%.

In the quest to develop preventive and therapeutic strategies, the first step has been to characterize disease progression itself. Over a century ago, Alois Alzheimer first identified a case of dementia for which post-mortem brain tissue demonstrated

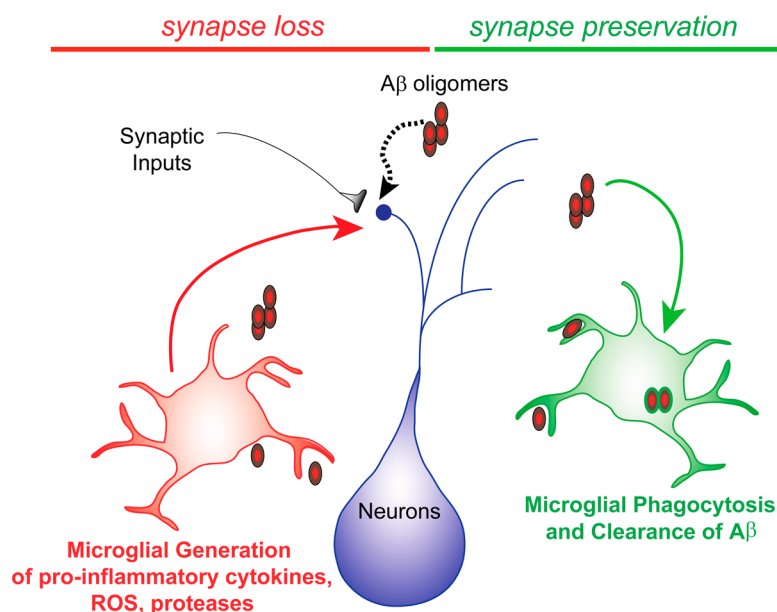
abnormal extracellular protein deposits and intracellular tangled structures.<sup>2</sup> Since that initial discovery, the amyloid- $\beta$  (A $\beta$ ) peptide has been identified as the major component of extracellular plaques in AD<sup>3</sup> and the microtubule associated protein tau as the major component of intracellular neurofibrillary tangles.<sup>4</sup> Supporting a causative role for A $\beta$  in AD, mutations in both amyloid precursor protein (APP) and presenilin-1 (PS1), which cleaves APP to form A $\beta$ , are associated with pedigrees in which early onset AD is genetically inherited.<sup>5,6</sup> These findings are the basis of the "amyloid hypothesis" that A $\beta$  generation is a driving force in the

**Special Issue:** Neuroinflammation

**Received:** January 16, 2016

**Accepted:** March 15, 2016

**Published:** March 15, 2016



**Figure 1.** The microglial neuroinflammatory response in AD. In AD, accumulating soluble  $A\beta$  species lead to synaptic loss through direct toxicity to synapses (black dashed arrow) and through maladaptive activation of microglia that produce damaging reactive oxygen species (ROS), cytokines, proteases, and chemokines. Alternatively, microglial activation can exert beneficial effects and maintain synaptic integrity and homeostasis by efficient clearance and phagocytosis of toxic misfolded proteins, such as oligomeric or fibrillar  $A\beta$  species, as well as removal of damaged synapses.

development of AD.<sup>7</sup> Lending additional support to this hypothesis, an APP mutation that reduces  $A\beta$  production and protects against the development of AD has recently been identified.<sup>8</sup> At the same time, the amyloid hypothesis of AD is not without controversy. The spatial progression of tau-containing neurofibrillary tangles correlates better with neuronal death and AD progression than do  $A\beta$  plaque deposits.<sup>9</sup> Moreover,  $A\beta$  plaques are present in the brains of many cognitively normal elderly patients, although some studies suggest that this represents a preclinical stage of AD.<sup>10,11</sup> To resolve this controversy, additional studies of  $A\beta$  and its downstream pathological effects will be necessary.

$A\beta$  exerts a number of toxic effects on neurons, resulting in synapse loss, neuronal network dysfunction, and ultimately neuron death, as reviewed in ref 12. Strategies aimed at reducing  $A\beta$  levels in the brain are thus a major focus of AD research. Unfortunately, several recent trials of monoclonal antibodies for  $A\beta$  have failed to alter cognitive outcomes in AD patients. This may be due to overall limitations of this strategy, including potential adverse effects of cerebral microhemorrhages and edema associated with perivascular amyloid deposits.<sup>13</sup> Hope remains for future studies focused on anti- $A\beta$  strategies as preventive interventions among patients at risk for AD,<sup>14</sup> as long as care is taken to avoid adverse effects. However, the need remains for additional targets that may be able to mitigate the detrimental effects of  $A\beta$  in the brain. One such target is the neuroinflammatory response to  $A\beta$ , a complex process that may be the third factor, along with  $A\beta$  and tau, driving AD pathogenesis (reviewed in ref 15). However, neuroinflammation exerts both toxic and protective effects in disease progression, so general strategies to reduce inflammation may not be as effective as more selective approaches targeting maladaptive inflammation. Identifying and developing targeted strategies to reduce toxic effects or increase protective effects remains a potential avenue for future preventive or therapeutic interventions in AD.

## ■ DETRIMENTAL AND BENEFICIAL EFFECTS OF NEUROINFLAMMATION IN AD

Activation of the brain's inflammatory response has long been recognized as a hallmark of AD pathology. Numerous features found in the AD brain suggest that a robust inflammatory response occurs either as a precursor to or as a consequence of  $A\beta$  plaque deposition and neurodegeneration. Many of these features resemble those seen in the innate immune response to pathogens or tissue injury: activated microglia, reactive astrocytes, complement proteins, cytokines, chemokines, and enzymes that generate reactive oxygen species are all elevated in the AD brain (reviewed in refs 16 and 17). Increasing evidence supports the idea that soluble  $A\beta$  oligomers or fibrils act directly on microglia, the resident phagocytes of the central nervous system, to drive this inflammatory process in AD (Figure 1). Microglia phagocytose  $A\beta$  in a manner dependent on scavenger receptors including CD36.<sup>18</sup> This uptake of  $A\beta$  activates inflammatory pathways, including those that lead to secretion of the inflammatory cytokine IL-1 $\beta$ .<sup>19</sup> In addition,  $A\beta$  promotes signaling via Toll-like receptors in microglia that drives production of reactive oxygen species and activation of the canonical pro-inflammatory transcription factor NF- $\kappa$ B.<sup>20</sup> Moreover, genome-wide analysis of microglia treated with  $A\beta$  shows enriched up-regulation of genes containing promoter sequences for NF- $\kappa$ B and IRF1/IRF7, other canonical mediators of inflammatory responses.<sup>21</sup>

In addition, genome-wide association study (GWAS) data from late-onset AD patients has identified variants in a number of genes implicated in the microglial inflammatory response. The E4 allele of the *APOE* gene is by far the most frequent genetic variant in late-onset AD and one of the earliest identified;<sup>22</sup> its role in the microglial inflammatory response is becoming increasingly clear. Microglia expressing the E4 allele of *APOE* have increased levels of pro-inflammatory COX-2 and prostaglandin production, with decreased levels of the microglial receptor TREM2.<sup>23</sup> Alleles of *TREM2* itself have also been

associated with AD.<sup>24</sup> The precise mechanism by which TREM2 contributes to AD is an active area of current study, with two recent reports indicating that TREM2 deficiency reduces reactive microgliosis in mouse models of AD, although the ultimate effect on A $\beta$  accumulation and neuronal survival appears to depend on the mouse model and time point examined.<sup>25,26</sup> Several other genetic polymorphisms associated with AD, including those in CD33 and complement receptor 1, have been found to play important roles in microglial inflammatory responses or clearance of A $\beta$  (reviewed in ref 15). Taken together, these studies suggest that impaired microglial function plays a central role in driving AD pathogenesis.

A major hypothesis of AD etiology poses that this ongoing inflammatory response promotes synaptotoxicity and other neuronal damage leading to AD. This idea is bolstered by a number of epidemiological studies showing that the use of nonsteroidal anti-inflammatory drugs (NSAIDs) is associated with reduced risk of AD, reviewed in refs 17 and 27, as discussed in more detail below. At the same time, other studies suggest that the brain inflammatory response may play a beneficial role, particularly with respect to the clearance of toxic A $\beta$  species, as reviewed in ref 28 and discussed below. A more complete understanding of the complex balance between toxic and beneficial roles of inflammation in AD will be necessary for improved approaches to AD prevention and treatment.

The microglial inflammatory response to A $\beta$  promotes a number of processes that appear to cause additional damage to neurons. Much of this evidence comes from mouse models of familial AD, where transgenes drive nervous system expression of mutant forms of the amyloid precursor protein (APP) or presenilin 1 (PS1) genes or both. For example, a recent study showed that microglial proliferation, which is increased in the AD patient brain, appears to drive synaptic toxicity. In APP<sub>Swe</sub>/PS1 $\Delta$ E9 transgenic mice, pharmacologically inhibiting microglial proliferation by blocking the CSF1 receptor leads to increased synaptic density and improved working memory behavior without altering A $\beta$  accumulation.<sup>29</sup> Specific molecular pathways within microglia appear to contribute to this neurotoxicity. C1q, the first component of the classical complement pathway, is elevated both in the AD brain and in the Tg2576 APP<sub>Swe</sub> transgenic mouse model. When C1q is genetically deleted, amyloid plaque levels remain largely constant while synaptic and dendritic loss is rescued, suggesting a role for complement in neuronal toxicity in AD.<sup>30</sup> Another inflammatory pathway, signaling through the receptor for advanced glycation endproducts (RAGE), appears to play an important role in promoting neuronal dysfunction. In the J9 APP<sub>Swe,Ind</sub> transgenic mouse model, genetic overexpression of RAGE accelerates spatial memory deficits in the radial arm water maze, while expression of a dominant negative form of RAGE rescues performance in the same task.<sup>31</sup> Taken together with the clear protective effects of NSAIDs in preventing AD,<sup>27</sup> these results suggest a role for inflammatory processes in accelerating neuronal damage and consequent memory deficits in mouse models of AD.

At the same time, inflammation may perform valuable functions such as clearance of toxic A $\beta$  peptides from the brain. Complement, for example, which appears to have toxic functions in some contexts,<sup>30</sup> may also perform valuable functions in promoting A $\beta$  clearance. Overexpression of the complement inhibitor protein sCrry in the J20 APP<sub>Swe,Ind</sub> model of AD results in reduced microglial activation, increased A $\beta$

plaque deposition, and loss of neurons in the CA3 region of the hippocampus.<sup>32</sup> The central importance of microglial clearance of A $\beta$  has been demonstrated similarly in a study examining the role of the cytokine TGF- $\beta$  in the J9 APP<sub>Swe,Ind</sub> model, where overexpression of TGF- $\beta$  increases microglial activation, reduces A $\beta$  deposition in the brain parenchyma, and reduces numbers of dystrophic neurites.<sup>33</sup> In addition to this role for cytokines in promoting the A $\beta$  clearance capacity of microglia, chemokine recruitment of microglia to sites of A $\beta$  accumulation appears to be another important beneficial pathway. The chemokine receptor CCR2, expressed by macrophages that infiltrate the nervous system after injury,<sup>34</sup> mediates migration of immune cells to sites of inflammation. In the Tg2576 APP<sub>Swe</sub> mouse model, genetic deletion of CCR2 results in reduced brain microglia and macrophage activation, increased brain A $\beta$  levels, and a dramatically accelerated mortality for both CCR2+/- and CCR2-/- Tg2576 mice.<sup>35</sup> These studies have in common the finding that lower levels of microglial activation are associated with higher A $\beta$  levels and worse outcomes at the neuronal or organismal level, suggesting that the microglial inflammatory response carries out important beneficial functions in AD. At the same time, it will be important for future research to note that what is observed as “microglial activation” (often by immunostaining for the expression of marker proteins) likely constitutes several different activation states that may play context-dependent toxic or beneficial roles.<sup>36</sup> Specifically, an age-dependent increase in toxic cytokine expression by microglia, accompanied by a decrease in receptors and enzymes associated with A $\beta$  clearance, may underlie some of the seemingly conflicting studies identifying either toxic or beneficial roles for the microglial inflammatory response in AD models.<sup>37</sup> Strategies aiming to tip the balance away from toxic effects and toward beneficial actions of microglia will thus be particularly important for translational studies in the future.

## ■ MIXED FINDINGS FOR NONSTEROIDAL ANTI-INFLAMMATORY DRUGS IN AD PREVENTION AND TREATMENT

Extensive epidemiological, clinical, and basic science literature supports a role of NSAIDs in preventing AD. The initial finding of this association came from McGeer and colleagues,<sup>38</sup> who reported a surprisingly low comorbidity for rheumatoid arthritis and AD, demonstrating a lower risk of AD in a patient population likely to take NSAIDs. Subsequent analysis of epidemiological data from several studies confirmed that chronic NSAID use was associated with lower risk of AD in normal aging populations.<sup>39</sup> Strikingly, prospective studies in targeted populations in Maryland,<sup>40</sup> Utah,<sup>41</sup> and The Netherlands,<sup>42</sup> and in the US Veteran's Affairs system<sup>43</sup> confirmed the protective effects of NSAIDs. Moreover, these studies found that longer NSAID use was associated with significantly greater protection, with up to 80% risk reduction for patients who had taken NSAIDs for more than two years.<sup>42</sup> These consistent findings present a compelling case for the exploration of NSAID-based mechanisms in the prevention of AD. However, as discussed below, the story of NSAID use as a protective strategy carries several major caveats.

Beyond the well-known gastric and renal complications of long-term NSAID use, close examination of epidemiological and clinical trial data suggests that if NSAIDs have a protective effect, it is only when started long before the onset of AD. Later in disease progression, NSAIDs may even worsen AD

Table 1. Selected Studies on NSAIDs in Mouse Models of AD

ref and year	model	drug used	age start (months)	treat time	age end (months)	A $\beta$ effect	inflamm. markers	neuronal or behavioral effect
55, 2000	Tg2576	ibuprofen	10	6 months	16	decreased	decreased	fewer dystrophic neurites
88, 2001	Tg2576	ibuprofen	10	6 months	16			rescued open field; lowered caspase activation
53, 2001	Tg2576	ibuprofen	14	17 months	17	decreased	decreased	
89, 2002	TG2576 x PS1M146L	ibuprofen	3	3 days	3	decreased		
		naproxen	3	3 days	3	decreased		
		ibuprofen	7	5 months	12	decreased	no change	
		NCX-2216	7	5 months	12	decreased	increased	
90, 2003	Tg2576	13 different NSAIDs	3	3 days	3	decreased		
91, 2003	Tg2576	indomethacin	12	8 months	20	decreased	no change	
57, 2003	Tg2576	ibuprofen	11	4 months	15	decreased	decreased	
92, 2004	Tg2576	pioglitazone	11	4 months	15	decreased	no change	
		indomethacin	8	7 months	15	decreased	decreased	
54, 2005	APPV717I	ibuprofen	10	7 days	10	decreased	decreased	
93, 2005	Tg2576	pioglitazone	10	7 days	10	decreased	decreased	
		flurbiprofen	3–4	3 days	3–4	decreased		
94, 2005	APP-PS1	flurbiprofen	8	6 months	14	no change	no change	
		NO-flurbiprofen	8	6 months	14	decreased	decreased	
95, 2007	Tg2576	flurbiprofen	8 or 17	6 months or 2 weeks	14 or 19	no change		rescued Morris water maze
60, 2008	Tg2576	ibuprofen	12	1 month	13	no change	decreased	rescued Morris water maze
		ibuprofen	5	5 months	10		decreased	rescued Morris water maze
		MF tricyclic	12	1 month	13	no change	decreased	rescued Morris water maze
		naproxen	12	1 month	13	no change	decreased	rescued Morris water maze
96, 2008	3xTg-AD	ibuprofen	1	5 months	6	decreased		rescued Morris water maze
61, 2009	R1.40	ibuprofen	3	3 months	6	no change	decreased	lowered markers of neuronal cell-cycle entry
		naproxen	3	3 months	6	no change	decreased	lowered markers of neuronal cell-cycle entry
56, 2012	R1.40	ibuprofen	15	9 months	24	decreased	decreased	
59, 2012	5x-FAD	ibuprofen	3	3 months	6	no change	decreased	worsened cross maze
97, 2013	3xTg-AD	flurbiprofen	5	2 months	7	no change		rescued radial arm water maze
98, 2013	3xTg-AD	SC-560	20	8 days	20	decreased	decreased	rescued Morris water maze
99, 2013	R1.40	tolfenamic acid	14–20	1 month	15–21	decreased		rescued Morris water maze and Y Maze
100, 2014	3xTg-AD	ibuprofen	1	15–22 months	16–23	decreased		rescued hippocampal volume by MRI

symptoms. Several major studies on NSAIDs and AD have noted that NSAID use, even for several years, does not confer protection if it occurs less than two years before AD onset.<sup>40–42</sup> Studies of normal age-related cognitive decline confirm this effect: in a population based study, patients who started NSAID use before age 65 demonstrated slower cognitive decline in their 70s and 80s, whereas those who started NSAID use after age 65 had either no change or accelerated cognitive decline depending on the population examined.<sup>44</sup> These observational findings provide hope for strategies based on NSAID use in middle age, many years before AD onset would normally occur.

Unfortunately, the clinical trials and randomized studies performed to date have generally focused on later stages of cognitive decline and have largely found no beneficial effect of NSAID treatment at this stage (reviewed in ref 17).

The general failure of these clinical trials for NSAIDs in AD reinforces the hypothesis that NSAID use is protective only if initiated long before disease onset. While one initial study found improved cognitive performance with indomethacin treatment compared with placebo in a small group of AD patients,<sup>45</sup> other trials have been disappointing. Neither naproxen nor rofecoxib had any effect on cognitive decline in



1-year placebo-controlled trials in patients with established AD.<sup>46,47</sup> In an attempt to study the effect of earlier NSAID use in nondemented populations, the AD Anti-Inflammatory Prevention Trial (ADAPT) was initiated to test in a placebo-controlled study whether naproxen or celecoxib prevented AD onset and slowed cognitive decline in patients over 70 years old at the time of enrollment. Unfortunately, treatment in this study was halted after cardiovascular complications were discovered with the use of these drugs in other clinical trials.<sup>48</sup> In ADAPT, treatment with either celecoxib or naproxen was terminated prematurely because of cardiovascular safety concerns; as a result, treatment with either celecoxib or naproxen lasted a median of 14.8 months, as opposed to the originally planned 7 years. Although a follow-up study on these subjects revealed no change in the risk of AD development 6 years after discontinuation of treatment,<sup>52</sup> other follow-up analyses have yielded additional insights. Analysis from the first four years after randomization demonstrated an increased risk for AD development among celecoxib- and naproxen-treated patients,<sup>49</sup> consistent with the worsening of cognitive decline seen in some populations who begin NSAID use after age 65.<sup>44</sup> Later reanalysis of the ADAPT study removed from the data set patients with any symptoms of cognitive impairment at the start of the trial and those who developed AD in the first 2.5 years of observation, thereby limiting the study to patients who were cognitively intact at the outset of the trial. In this post hoc analysis, naproxen-treated patients showed significantly reduced rates of AD.<sup>50</sup> Moreover, CSF samples taken from a randomized subset of these patients demonstrated that naproxen significantly reduced the tau/A $\beta$ 42 ratio, a biomarker positively correlated with the risk of cognitive decline and AD among elderly populations.<sup>51</sup> While these data come from a post hoc analysis of a trial initially designed for different measures, they offer a hint of support for the preventive role of NSAIDs in AD.

One possibility is that long-term NSAID use protects against AD only when initiated long before cognitive decline begins. The failure of previous clinical trials in AD patients likely reflects the paradoxically detrimental effects of NSAID use when cognitive decline leading to AD has already begun. Future clinical studies will thus need to examine the longer-term effects of NSAID treatment beginning in younger patients. These trials, however, will be challenging to pursue both in terms of time and funding. Moreover, the side effects associated with NSAID use suggest that identifying the downstream mechanism for NSAID protection could yield safer, more targeted interventions. Translational studies to identify these mechanisms in animal models may thus offer key insight for AD prevention strategies.

To date, more than 20 studies have assessed the effects of NSAID treatment in transgenic mouse models overexpressing mutant APP or PS1 or both, as listed in Table 1. Many of these studies have focused on the ability of NSAIDs to modulate brain A $\beta$  levels, because NSAIDs such as ibuprofen modulate  $\gamma$ -secretase activity to produce less A $\beta$ 42 from APP in cultured cells.<sup>53</sup> In general, the studies that have administered NSAIDs to mice after the onset of amyloid plaque deposition<sup>54–57</sup> have found reduced amyloid plaque load and microglial activation in the brains of AD model mice treated with NSAIDs such as ibuprofen over the course of several months. However, it is important to note that this effect depends on which PS1 mutant is used in the model, because many familial PS1 mutations render the protein insensitive to  $\gamma$ -secretase modification by

NSAIDs.<sup>58</sup> Indeed, ibuprofen treatment reduces inflammatory markers but has no effect on A $\beta$  levels or working memory in the 5x-FAD model containing one such mutant form of PS1.<sup>59</sup> The above data are consistent with the proposed A $\beta$ -lowering mechanism for NSAID protection against AD. Yet other studies have found that NSAIDs can reverse spatial memory deficits in Tg2576 APP<sub>Swe</sub> mice without altering A $\beta$  levels or markers of inflammation,<sup>60</sup> calling in to question the hypothesis that NSAIDs achieve protection through reduced A $\beta$ 42 generation or even through anti-inflammatory effects. Moreover, ibuprofen treatment before plaque onset in the slow-progressing R1.40 model prevents the onset of neuronal cell cycle re-entry, a marker of vulnerable neurons in AD, without altering APP processing or A $\beta$  levels.<sup>61</sup> These conflicting results suggest the need for animal studies that mirror the effects of NSAIDs in human patients as much as possible to uncover the mechanisms at work in the reduced risk of AD. In addition, the molecular targets and downstream signaling effects of NSAIDs may yield more specific answers.

## ■ DETRIMENTAL AND BENEFICIAL EFFECTS OF PROSTAGLANDIN SIGNALING IN AD

NSAIDs inhibit cyclooxygenase enzymes—the constitutively expressed COX-1 and the inducible COX-2, cytosolic enzymes that generate PGH<sub>2</sub> from membrane stores of arachidonic acid. PGH<sub>2</sub> is further modified to produce the prostaglandins PGE<sub>2</sub>, PGD<sub>2</sub>, PGI<sub>2</sub>, and PGF<sub>2 $\alpha$</sub> , and the thromboxane TXA<sub>2</sub>. While the effect of NSAIDs to oppose inflammation has led to a generalized view of COX activity as toxic in neuroinflammatory disease, recent studies demonstrate a diversity of both toxic and protective functions for the prostaglandin products of COX-1 and COX-2. This divergence became vividly clear in the clinical development and subsequent discontinuation of several selective COX-2 inhibitors, originally aimed at inhibiting toxic COX-2 activity while avoiding inhibition of protective COX-1 activity in the gut. More extensive clinical studies found an unexpected increase in the rates of myocardial infarction and thrombotic stroke among patients on COX-2 inhibitors; however, this began only after 18 months of treatment.<sup>48,62</sup> One hypothesized mechanism for the toxic effect of COX-2 inhibition lies in the balance of COX-2 derived prostacyclin, which promotes vasodilation and restrains platelet activation, and COX-1 derived thromboxane, which promotes vasoconstriction and aggregation of platelets.<sup>63,64</sup> Another potential mechanism may involve an off-target effect of compounds that manifests very late after chronic COX-2 suppression. This finding indicates that, while some prostaglandin pathways are toxic, others are beneficial in a manner that is likely context dependent. Moreover, the general reluctance to use COX-2 inhibitors in the clinic underscores the need for identification of more specific interventions downstream of COX activity.

To this end, a number of recent studies have investigated the role of the downstream prostaglandin products and their receptors in models of neurological disease. Among the prostaglandins, PGE<sub>2</sub> is of particular interest because it is found at relatively high concentrations in the brain.

PGE<sub>2</sub> levels are increased in the CSF of probable AD patients<sup>65</sup> and in AD patients with mild memory impairment.<sup>66</sup> PGE<sub>2</sub> exerts its cellular effects through four distinct G protein-coupled E-prostanoid receptors, EP1 to EP4. In vivo localization of PGE<sub>2</sub> receptors in the brain has been limited because of poor specificity and high background of available antibodies to EP receptors; however in situ hybridization studies of mouse

brain indicate that the EP1 and to a lesser extent the EP3 and EP4 receptors are expressed basally in neurons of forebrain and cerebellum (Allen Brain Atlas, [www.alleninstitute.org](http://www.alleninstitute.org)). As described in detail below, in settings of innate immune neuroinflammation, expression levels of EP2 and EP4 transcripts are induced in microglia. Activation of PGE<sub>2</sub> receptors triggers intracellular signals that lead to modifications in production of cAMP or phosphoinositol (PI) turnover. The effects of PGE<sub>2</sub> are complex because PGE<sub>2</sub> can bind to four distinct EP (for E-prostanoid) receptors, EP1 through EP4, that have divergent second messenger systems. The EP2 and EP4 receptors are coupled to G<sub>αs</sub>, increasing cAMP production upon PGE<sub>2</sub> binding; EP3 is coupled to G<sub>αi</sub> and decreases cAMP production; while EP1 is coupled to G<sub>αq</sub> and increases intracellular Ca<sup>2+</sup> concentrations.<sup>67</sup> Consistent with findings of beneficial as well as toxic effects of COX activity, emerging evidence indicates both protective and toxic functions for the EP receptors in ways that are receptor-, context-, and cell-type-specific, as reviewed in refs 68 and 69. However, a growing number of studies on chronic inflammatory models suggest that EP1, EP2, and EP3 receptors promote neuronal injury in models of chronic neurodegenerative disease, whereas EP4 exerts largely beneficial effects.

Of the four EP receptors, the role of EP1 in inflammatory neurodegenerative disease is least clear. EP1 exerts neurotoxic effects in models of oxidative stress, where inhibition of EP1 signaling rescues dopaminergic neurons from 6-hydroxydopamine mediated cell death.<sup>70</sup> EP1 also appears to play a role in models of AD: genetic deletion of EP1 in the APP<sub>Swe</sub>-PS1<sub>ΔE9</sub> mouse model of AD reduces amyloid plaque numbers in the hippocampus.<sup>71</sup> However, the specific role of EP1 in microglia or other cell types remains to be specified.

EP2, by contrast, has been widely studied in models of inflammatory neurodegeneration. Deletion of EP2 in the APP<sub>Swe</sub>/PS1<sub>ΔE9</sub> mouse model of AD results in decreased brain lipid oxidation and reduced A $\beta$  levels, potentially through reduced activity of the APP cleaving enzyme BACE-1,<sup>72</sup> an enzyme whose expression and enzymatic activity is responsive to oxidative damage in neurons.<sup>73</sup> The functions of EP2 in promoting oxidative damage extend more generally to models of LPS-induced innate immune activation, where EP2 deletion completely abolishes the increase in brain lipid oxidation after immune activation.<sup>74</sup> Here, at least some of the toxicity of EP2 is due to activity in microglia: cultured microglia from EP2 deficient mice fail to mount a neurotoxic oxidative response to LPS.<sup>75</sup> Moreover, EP2 is a central player in the neurotoxic response of microglia to A $\beta$ : as in the case of LPS, cultured microglia exposed to A $\beta$  promote neuron death, but not when the microglia are deficient for EP2. Here, EP2 may promote further damage in models of AD, because signaling through EP2 suppresses the ability of microglia to phagocytose A $\beta$ .<sup>76</sup> Consistent with this finding, lethally irradiated APP<sub>Swe</sub>/PS1<sub>ΔE9</sub> mice show reduced cerebral A $\beta$  burden when their microglial populations are reconstituted with EP2-deficient bone marrow-derived cells compared with wild-type cells.<sup>77</sup> The mechanism behind this effect may lie at least partially in the ability of EP2 signaling to suppress expression of the scavenger receptor CD36, a key mediator of A $\beta$  phagocytosis by microglia.<sup>78</sup> To specifically assess the role of microglial EP2 in models of AD, recent studies using the Cre/loxP system to delete EP2 in microglia<sup>79</sup> have shown increased brain insulin-like growth factor levels, restored working memory, and increased synaptic density in APP<sub>Swe</sub>/PS1<sub>ΔE9</sub> mice lacking EP2 specifically in

microglia.<sup>80</sup> Thus, microglial EP2 may promote toxicity in AD through the multiple mechanisms of reducing A $\beta$  clearance, increasing the microglial oxidative response to A $\beta$ , and suppressing protective growth factor production.

The EP3 receptor, though less studied so far in models of AD, appears to possess similar properties to the EP2 receptor. Deletion of EP3 in APP<sub>Swe</sub>/PS1<sub>ΔE9</sub> mice reduces inflammatory protein expression, markers of oxidative stress, A $\beta$  levels, and BACE-1 activity.<sup>81</sup> Here again, the response of BACE-1 to oxidative damage<sup>73</sup> may underlie some of the anti-amyloidogenic effects of EP3 deletion, or EP3 may regulate the clearance of A $\beta$  by microglia. These studies suggest that, although EP1, EP2, and EP3 receptors signal through distinct and even opposing G-protein coupled pathways, they each contribute to oxidative stress and damaging inflammatory responses in chronic models of AD.

In contrast, the EP4 receptor appears to exert protective effects in models of innate immunity and AD, as well as neuroprotective effects against hypoxic damage in models of stroke.<sup>82</sup> As detailed above, stimulation of the innate immune response through peripheral injection of LPS in mice elicits a brain inflammatory response characterized by microglial activation, cytokine secretion, and production of reactive oxygen and nitrogen species that increase brain lipid oxidation. Direct stimulation of EP4 receptors in cultured microglia dramatically reduces this response to LPS,<sup>83,84</sup> suggesting that EP4 signaling is sufficient for attenuation of the microglial innate immune response. The substrate for this divergence between EP2 and EP4 inflammatory signaling may lie in functions encoded in the extended carboxy-tail of EP4, which binds proteins that signal to inhibit inflammatory NF- $\kappa$ B transcriptional activity.<sup>83</sup> In line with this, EP4 appears to be necessary for resolution of the inflammatory response. In wild-type mice, cytokine and oxidative enzyme expression in the brain is significantly elevated by 6 h after LPS injection but largely returns to baseline by 24 h. In mice with selective deletion of EP4 in microglia, this inflammatory response in the brain persists at least to 24 h.<sup>84</sup> The anti-inflammatory effect of EP4 signaling appears to extend to the periphery as well: genetic deletion or pharmacological inhibition of EP4 exacerbates inflammation and tissue damage in a model of ulcerative colitis.<sup>85</sup> These findings strongly implicate EP4 signaling as a pivotal factor in resolving the innate immune response in inflammatory disease. Recent studies have extended this to models of AD as well: pharmacological stimulation of the EP4 receptor in cultured microglia reduces the inflammatory response to A $\beta$  and promotes phagocytic clearance of A $\beta$ , while conditional deletion of the EP4 receptor in microglia exacerbates inflammatory responses, A $\beta$  accumulation, and synaptic protein loss in APP<sub>Swe</sub>/PS1<sub>ΔE9</sub> mice.<sup>21</sup> Taken together, these data support an anti-inflammatory and protective role for microglial EP4 signaling in models of AD.

The above data suggest a complex mechanism in which signaling by one ligand, PGE<sub>2</sub>, may promote both an initial inflammatory response (e.g., through EP2 or EP3) and a compensatory resolution of immune activation through EP4. Given the findings above, several potential therapeutic strategies emerge to combat the damaging inflammatory response in AD: suppressing the toxic effects of EP1, EP2, and EP3 signaling or harnessing the protective effects of EP4 signaling. In the AD brain in particular, the balance among EP receptors appears to be weighted toward maladaptive pro-inflammatory signaling, as EP3 protein levels increase while

EP4 protein levels decrease in the brains of mild cognitive impairment (MCI) and AD patients.<sup>21,81</sup> This mechanism also presents a potential explanation for the contrasting protective and detrimental effects of NSAIDs seen in studies of patients at different stages of disease progression. It may be that inhibition of prostaglandin production is beneficial only at early stages when it can adequately suppress toxic signaling through EP2 and EP3 while preserving beneficial signaling through EP4.

As an additional layer of complexity, COX-1, COX-2, and EP receptors are expressed in other cell types including neurons, where their effects and contributions to AD progression may be distinct from those in microglia. COX-2 itself is a neuronal immediate early gene whose expression is rapidly induced in response to NMDA-dependent synaptic activity in the cortex and hippocampus.<sup>86</sup> Excessive neuronal COX-2 activity may be another contributing factor to AD, because neuronal overexpression of COX-2 in the APP<sub>Swe</sub>/PS1<sub>ΔE9</sub> mouse model worsens spatial working memory behavior.<sup>87</sup> In light of this, neuronal COX-2 may be yet another target of NSAIDs that could contribute to their protective effects, in addition to their suppression of damaging pathways in the neuroinflammatory response.

### ■ CONCLUSION: THE NEED FOR MOLECULAR, CELLULAR, AND TEMPORAL SPECIFICITY IN AD RESEARCH

In light of the clinical and animal model studies discussed above, there may be limited value in pursuing interventions aimed at broad-scale inhibition of COX activity in AD without regard to timing, cellular specificity, or downstream signaling complexity. The mixed evidence for preventive effects of NSAIDs in AD and the contrasting effects of NSAIDs in different mouse models at different pathological stages accentuate the need to identify which mechanism(s) underlie NSAID protection so that more targeted strategies can be pursued. Within prostaglandin signaling, the balance between protective and toxic signaling through different PGE<sub>2</sub> receptors underscores the need for additional detailed investigations of cell- and receptor-specific strategies. In the wider neuroinflammatory response to A $\beta$ , these studies add to a growing consensus that it is too simplistic to think of the inflammatory response in AD as entirely damaging or entirely protective. Instead, future studies targeting individual components of the neuroinflammatory response may yield new insights in the search for preventive and therapeutic strategies against AD.

### ■ AUTHOR INFORMATION

#### Corresponding Author

\*K. Andreasson. E-mail: [kandreas@stanford.edu](mailto:kandreas@stanford.edu).

#### Present Address

†N.S.W.: Institute of Healthy Aging, University College London, London, U.K.

#### Notes

The authors declare no competing financial interest.

### ■ ACKNOWLEDGMENTS

Authors supported by RO1AG030209 (KIA), R01AG15799 (KIA), R21AG033914 (KIA), RO1AG048232 (KIA), P50 AG047366 (KIA), Alzheimer's Association (KIA), Bright Focus (KIA), and NRSA F31AG039195 (NSW).

### ■ REFERENCES

- (1) Hebert, L. E., Scherr, P. A., Bienias, J. L., Bennett, D. A., and Evans, D. A. (2003) Alzheimer disease in the US population: prevalence estimates using the 2000 census. *Arch. Neurol.* **60**, 1119–1122.
- (2) Alzheimer, A. (1907) Über eine eigenartige Erkrankung der Hirnrinde [About a peculiar disease of the cerebral cortex]. *Allg. Z. Psychiatr. Psych.-Gerichtl. Med.* **64**, 146–148.
- (3) Glenner, G. G., and Wong, C. W. (1984) Alzheimer's disease: initial report of the purification and characterization of a novel cerebrovascular amyloid protein. *Biochem. Biophys. Res. Commun.* **120**, 885–890.
- (4) Grundke-Iqbal, I., Iqbal, K., Tung, Y. C., Quinlan, M., Wisniewski, H. M., and Binder, L. I. (1986) Abnormal phosphorylation of the microtubule-associated protein tau (tau) in Alzheimer cytoskeletal pathology. *Proc. Natl. Acad. Sci. U. S. A.* **83**, 4913–4917.
- (5) Goate, A., Chartier-Harlin, M. C., Mullan, M., Brown, J., Crawford, F., Fidani, L., Giuffra, L., Haynes, A., Irving, N., James, L., et al. (1991) Segregation of a missense mutation in the amyloid precursor protein gene with familial Alzheimer's disease. *Nature* **349**, 704–706.
- (6) Sherrington, R., Rogae, E. I., Liang, Y., Rogae, E. A., Levesque, G., Ikeda, M., Chi, H., Lin, C., Li, G., Holman, K., Tsuda, T., Mar, L., Foncin, J. F., Bruni, A. C., Montesi, M. P., Sorbi, S., Rainero, I., Pinessi, L., Nee, L., Chumakov, I., Pollen, D., Brookes, A., Sanseau, P., Polinsky, R. J., Wasco, W., Da Silva, H. A., Haines, J. L., Pericak-Vance, M. A., Tanzi, R. E., Roses, A. D., Fraser, P. E., Rommens, J. M., and St George-Hyslop, P. H. (1995) Cloning of a gene bearing missense mutations in early-onset familial Alzheimer's disease. *Nature* **375**, 754–760.
- (7) Hardy, J. A., and Higgins, G. A. (1992) Alzheimer's disease: the amyloid cascade hypothesis. *Science* **256**, 184–185.
- (8) Jonsson, T., Atwal, J. K., Steinberg, S., Snaedal, J., Jonsson, P. V., Björnsson, S., Stefansson, H., Sulem, P., Gudbjartsson, D., Maloney, J., Hoyte, K., Gustafson, A., Liu, Y., Lu, Y., Bhargale, T., Graham, R. R., Huttenlocher, J., Björnsdóttir, G., Andreassen, O. A., Jönsson, E. G., Palotie, A., Behrens, T. W., Magnusson, O. T., Kong, A., Thorsteinsdóttir, U., Watts, R. J., and Stefansson, K. (2012) A mutation in APP protects against Alzheimer's disease and age-related cognitive decline. *Nature* **488**, 96–99.
- (9) Arriagada, P. V., Growdon, J. H., Hedley-Whyte, E. T., and Hyman, B. T. (1992) Neurofibrillary tangles but not senile plaques parallel duration and severity of Alzheimer's disease. *Neurology* **42**, 631–631.
- (10) Morris, J. C., Storandt, M., McKeel, D. W., Rubin, E. H., Price, J. L., Grant, E. A., and Berg, L. (1996) Cerebral amyloid deposition and diffuse plaques in normal aging: Evidence for presymptomatic and very mild Alzheimer's disease. *Neurology* **46**, 707–719.
- (11) Vlassenko, A. G., Mintun, M. A., Xiong, C., Sheline, Y. I., Goate, A. M., Benzinger, T. L. S., and Morris, J. C. (2011) Amyloid-beta plaque growth in cognitively normal adults: longitudinal [11C]-Pittsburgh compound B data. *Ann. Neurol.* **70**, 857–861.
- (12) Palop, J. J., and Mucke, L. (2010) Amyloid-beta-induced neuronal dysfunction in Alzheimer's disease: from synapses toward neural networks. *Nat. Neurosci.* **13**, 812–818.
- (13) Wisniewski, T., and Konietzko, U. (2008) Amyloid-beta immunisation for Alzheimer's disease. *Lancet Neurol.* **7**, 805–811.
- (14) Miller, G. (2012) Alzheimer's research. Stopping Alzheimer's before it starts. *Science* **337**, 790–792.
- (15) Mhatre, S. D., Tsai, C. A., Rubin, A. J., James, M. L., and Andreasson, K. I. (2015) Microglial Malfunction: The Third Rail in the Development of Alzheimer's Disease. *Trends Neurosci.* **38**, 621–636.
- (16) Akiyama, H. H., Barger, S. S., Barnum, S. S., Bradt, B. B., Bauer, J. J., Cole, G. M. G., Cooper, N. R. N., Eikelenboom, P. P., Emmerling, M. M., Fiebich, B. L. B., Finch, C. E. C., Frautschy, S. S., Griffin, W. S. W., Hampel, H. H., Hull, M. M., Landreth, G. G., Lue, L. L., Mrak, R. R., Mackenzie, I. R. I., McGeer, P. L. P., O'Banion, M. K. M., Pachter, J. J., Pasinetti, G. G., Plata-Salaman, C. C., Rogers, J. J., Rydel, R. R.,



- Shen, Y. Y., Streit, W. W., Strohmeyer, R. R., Tooyoma, I. I., Van Muiswinkel, F. L. F., Veerhuis, R. R., Walker, D. D., Webster, S. S., Wegryniak, B. B., Wenk, G. G., and Wyss-Coray, T. T. (2000) Inflammation and Alzheimer's disease. *Neurobiol. Aging* 21, 383–421.
- (17) Heneka, M. T., Carson, M. J., Khoury, J. E., Landreth, G. E., Brosseron, F., Feinstein, D. L., Jacobs, A. H., Wyss-Coray, T., Vitorica, J., Ransohoff, R. M., Herrup, K., Frautschy, S. A., Finsen, B., Brown, G. C., Verkhratsky, A., Yamanaka, K., Koistinaho, J., Latz, E., Halle, A., Petzold, G. C., Town, T., Morgan, D., Shinohara, M. L., Perry, V. H., Holmes, C., Bazan, N. G., Brooks, D. J., Hunot, S., Joseph, B., Deigendesch, N., Garaschuk, O., Boddeke, E., Dinarello, C. A., Breitner, J. C., Cole, G. M., Golenbock, D. T., and Kummer, M. P. (2015) Neuroinflammation in Alzheimer's disease. *Lancet Neurol.* 14, 388–405.
- (18) El Khoury, J. B., Moore, K. J., Means, T. K., Leung, J., Terada, K., Toft, M., Freeman, M. W., and Luster, A. D. (2003) CD36 mediates the innate host response to beta-amyloid. *J. Exp. Med.* 197, 1657–1666.
- (19) Halle, A., Hornung, V., Petzold, G. C., Stewart, C. R., Monks, B. G., Reinheckel, T., Fitzgerald, K. A., Latz, E., Moore, K. J., and Golenbock, D. T. (2008) The NALP3 inflammasome is involved in the innate immune response to amyloid-beta. *Nat. Immunol.* 9, 857–865.
- (20) Reed-Geaghan, E. G., Savage, J. C., Hise, A. G., and Landreth, G. E. (2009) CD14 and Toll-Like Receptors 2 and 4 Are Required for Fibrillar A $\beta$ -Stimulated Microglial Activation. *J. Neurosci.* 29, 11982–11992.
- (21) Woodling, N. S., Wang, Q., Priyam, P. G., Larkin, P., Shi, J., Johansson, J. U., Zagol-Ikapitte, I., Boutaud, O., and Andreasson, K. I. (2014) Suppression of Alzheimer-Associated Inflammation by Microglial Prostaglandin-E2 EP4 Receptor Signaling. *J. Neurosci.* 34, 5882–5894.
- (22) Poirier, J., Davignon, J., Bouthillier, D., Kogan, S., Bertrand, P., and Gauthier, S. (1993) Apolipoprotein E polymorphism and Alzheimer's disease. *Lancet* 342, 697–699.
- (23) Li, X., Montine, K. S., Keene, C. D., and Montine, T. J. (2015) Different mechanisms of apolipoprotein E isoform-dependent modulation of prostaglandin E2 production and triggering receptor expressed on myeloid cells 2 (TREM2) expression after innate immune activation of microglia. *FASEB J.* 29, 1754–1762.
- (24) Guerreiro, R., Wojtas, A., Bras, J., Carrasquillo, M., Rogava, E., Majounie, E., Cruchaga, C., Sassi, C., Kauwe, J. S. K., Younkin, S., Hazrati, L., Collinge, J., Pockock, J., Lashley, T., Williams, J., Lambert, J.-C., Amouyel, P., Goate, A., Rademakers, R., Morgan, K., Powell, J., St George-Hyslop, P., Singleton, A., and Hardy, J. (2013) TREM2 Variants in Alzheimer's Disease. *N. Engl. J. Med.* 368, 117.
- (25) Jay, T. R., Miller, C. M., Cheng, P. J., Graham, L. C., Bemiller, S., Broihier, M. L., Xu, G., Margevicius, D., Karlo, J. C., Sousa, G. L., Coteleur, A. C., Butovsky, O., Bekris, L., Staugaitis, S. M., Leverenz, J. B., Pimplikar, S. W., Landreth, G. E., Howell, G. R., Ransohoff, R. M., and Lamb, B. T. (2015) TREM2 deficiency eliminates TREM2+ inflammatory macrophages and ameliorates pathology in Alzheimer's disease mouse models. *J. Exp. Med.* 212, 287–295.
- (26) Wang, Y., Cella, M., Mallinson, K., Ulrich, J. D., Young, K. L., et al. (2015) TREM2 Lipid Sensing Sustains the Microglial Response in an Alzheimer's Disease Model. *Cell* 160, 1061.
- (27) McGeer, P. L., and McGeer, E. G. (2007) NSAIDs and Alzheimer disease: epidemiological, animal model and clinical studies. *Neurobiol. Aging* 28, 639–647.
- (28) Wyss-Coray, T. (2006) Inflammation in Alzheimer disease: driving force, bystander or beneficial response? *Nat. Med.* 12, 1005–1015.
- (29) Olmos-Alonso, A., Schettters, S. T. T., Sri, S., Askew, K., Mancuso, R., Vargas-Caballero, M., Holscher, C., Perry, V. H., and Gomez-Nicola, D. (2016) Pharmacological targeting of CSF1R inhibits microglial proliferation and prevents the progression of Alzheimer's-like pathology. *Brain* 139, 891.
- (30) Fonseca, M. I., Zhou, J., Botto, M., and Tenner, A. J. (2004) Absence of C1q leads to less neuropathology in transgenic mouse models of Alzheimer's disease. *J. Neurosci.* 24, 6457–6465.
- (31) Arancio, O., Zhang, H. P., Chen, X., Lin, C., Trinchese, F., Puzzo, D., Liu, S., Hegde, A., Yan, S. F., Stern, A., Luddy, J. S., Lue, L.-F., Walker, D. G., Roher, A., Buttini, M., Mucke, L., Li, W., Schmidt, A. M., Kindy, M., Hyslop, P. A., Stern, D. M., and Du Yan, S. S. (2004) RAGE potentiates Abeta-induced perturbation of neuronal function in transgenic mice. *EMBO J.* 23, 4096–4105.
- (32) Wyss-Coray, T., Yan, F., Lin, A., Lambris, J., Alexander, J., Quigg, R., and Masliah, E. (2002) Prominent neurodegeneration and increased plaque formation in complement-inhibited Alzheimer's mice. *Proc. Natl. Acad. Sci. U. S. A.* 99, 10837–10842.
- (33) Wyss-Coray, T., Lin, C., Yan, F., Yu, G. Q., Rohde, M., McConlogue, L., Masliah, E., and Mucke, L. (2001) TGF-beta1 promotes microglial amyloid-beta clearance and reduces plaque burden in transgenic mice. *Nat. Med.* 7, 612–618.
- (34) Mizutani, M., Pino, P. A., Saederup, N., Charo, I. F., Ransohoff, R. M., and Cardona, A. E. (2012) The Fractalkine Receptor but Not CCR2 Is Present on Microglia from Embryonic Development throughout Adulthood. *J. Immunol.* 188, 29–36.
- (35) El Khoury, J., Toft, M., Hickman, S. E., Means, T. K., Terada, K., Geula, C., and Luster, A. D. (2007) Ccr2 deficiency impairs microglial accumulation and accelerates progression of Alzheimer-like disease. *Nat. Med.* 13, 432–438.
- (36) Colton, C. A. (2009) Heterogeneity of Microglial Activation in the Innate Immune Response in the Brain. *J. Neuroimmune Pharmacol* 4, 399–418.
- (37) Hickman, S. E., Allison, E. K., and El Khoury, J. (2008) Microglial Dysfunction and Defective Beta-Amyloid Clearance Pathways in Aging Alzheimer's Disease Mice. *J. Neurosci.* 28, 8354–8360.
- (38) McGeer, P. L., McGeer, E., Rogers, J., and Sibley, J. (1990) Anti-inflammatory drugs and Alzheimer disease. *Lancet* 335, 1037.
- (39) McGeer, P. L., Schulzer, M., and McGeer, E. G. (1996) Arthritis and anti-inflammatory agents as possible protective factors for Alzheimer's disease: A review of 17 epidemiologic studies. *Neurology* 47, 425–432.
- (40) Stewart, W. F., Kawas, C., Corrada, M., and Metter, E. J. (1997) Risk of Alzheimer's disease and duration of NSAID use. *Neurology* 48, 626–632.
- (41) Zandi, P. P., Anthony, J. C., Hayden, K. M., Mehta, K., Mayer, L., and Breitner, J. (2002) Reduced incidence of AD with NSAID but not H2 receptor antagonists: the Cache County Study. *Neurology* 59, 880–886.
- (42) in t Veld, B. A., Ruitenber, A., Hofman, A., Launer, L. J., van Duijn, C. M., Stijnen, T., Breteler, M. M., and Stricker, B. H. (2001) Nonsteroidal antiinflammatory drugs and the risk of Alzheimer's disease. *N. Engl. J. Med.* 345, 1515–1521.
- (43) Vlad, S. C., Miller, D. R., Kowall, N. W., and Felson, D. T. (2008) Protective effects of NSAIDs on the development of Alzheimer disease. *Neurology* 70, 1672–1677.
- (44) Hayden, K. M., Zandi, P. P., Khachaturian, A. S., Szekely, C. A., Fotuhi, M., Norton, M. C., Tschanz, J. T., Pieper, C. F., Corcoran, C., Lyketsos, C. G., Breitner, J. C. S., Welsh-Bohmer, K. A., and the Cache County Investigators (2007) Does NSAID use modify cognitive trajectories in the elderly? The Cache County study. *Neurology* 69, 275–282.
- (45) Rogers, J., Kirby, L. C., Hempelman, S. R., Berry, D. L., McGeer, P. L., Kaszniak, A. W., Zalinski, J., Cofield, M., Mansukhani, L., Willson, P., and Kogan, F. (1993) Clinical trial of indomethacin in Alzheimer's disease. *Neurology* 43, 1609–1611.
- (46) Aisen, P. S., Schafer, K. A., Grundman, M., Pfeiffer, E., Sano, M., Davis, K. L., Farlow, M. R., Jin, S., Thomas, R. G., Thal, L. J., and the Alzheimer's Disease Cooperative Study (2003) Effects of rofecoxib or naproxen vs placebo on Alzheimer disease progression: a randomized controlled trial. *JAMA, J. Am. Med. Assoc.* 289, 2819–2826.
- (47) Reines, S. A., Block, G. A., Morris, J. C., Liu, G., Nessly, M. L., Lines, C. R., Norman, B. A., Baranak, C. C., and the Rofecoxib Protocol 091 Study Group (2004) Rofecoxib: no effect on Alzheimer's



disease in a 1-year, randomized, blinded, controlled study. *Neurology* 62, 66–71.

(48) Bresalier, R. S., Sandler, R. S., Quan, H., Bolognese, J. A., Oxenius, B., Horgan, K., Lines, C., Riddell, R., Morton, D., Lanas, A., Konstam, M. A., Baron, J. A., and the Adenomatous Polyp Prevention on Vioxx (APPROVe) Trial Investigators (2005) Cardiovascular events associated with rofecoxib in a colorectal adenoma chemoprevention trial. *N. Engl. J. Med.* 352, 1092–1102.

(49) ADAPT Research Group, Martin, B. K., Szekely, C., Brandt, J., Piantadosi, S., Breitner, J. C. S., Craft, S., Evans, D., Green, R., and Mullan, M. (2008) Cognitive function over time in the Alzheimer's Disease Anti-inflammatory Prevention Trial (ADAPT): results of a randomized, controlled trial of naproxen and celecoxib. *Arch. Neurol.* 65, 896–905.

(50) Breitner, J. C., Baker, L. D., Montine, T. J., Meinert, C. L., Lyketsos, C. G., Ashe, K. H., Brandt, J., Craft, S., Evans, D. E., Green, R. C., Ismail, M. S., Martin, B. K., Mullan, M. J., Sabbagh, M., Tariot, P. N., and ADAPT Research Group (2011) Extended results of the Alzheimer's disease anti-inflammatory prevention trial. *Alzheimer's Dementia* 7, 402–411.

(51) Sonnen, J. A., Montine, K. S., Quinn, J. F., Kaye, J. A., Breitner, J. C., and Montine, T. J. (2008) Biomarkers for cognitive impairment and dementia in elderly people. *Lancet Neurol.* 7, 704–714.

(52) ADAPT Research Group (2013) Results of a follow-up study to the randomized Alzheimer's Disease Anti-inflammatory Prevention Trial (ADAPT). *Alzheimer's Dementia* 9, 714–723.

(53) Weggen, S., Eriksen, J. L., Das, P., Sagi, S. A., Wang, R., Pietrzik, C. U., Findlay, K. A., Smith, T. E., Murphy, M. P., Bulter, T., Kang, D. E., Marquez-Sterling, N., Golde, T. E., and Koo, E. H. (2001) A subset of NSAIDs lower amyloidogenic Abeta42 independently of cyclooxygenase activity. *Nature* 414, 212–216.

(54) Heneka, M. T., Sastre, M., Dumitrescu-Ozimek, L., Hanke, A., Dewachter, I., Kuiperi, C., O'Banion, K., Klockgether, T., van Leuven, F., and Landreth, G. E. (2005) Acute treatment with the PPARgamma agonist pioglitazone and ibuprofen reduces glial inflammation and Abeta1–42 levels in APPV717I transgenic mice. *Brain* 128, 1442–1453.

(55) Lim, G. P., Yang, F., Chu, T., Chen, P., Beech, W., Teter, B., Tran, T., Ubeda, O., Ashe, K. H., Frautschy, S. A., and Cole, G. M. (2000) Ibuprofen suppresses plaque pathology and inflammation in a mouse model for Alzheimer's disease. *J. Neurosci.* 20, 5709–5714.

(56) Wilkinson, B. L., Cramer, P. E., Varvel, N. H., Reed-Geaghan, E., Jiang, Q., Szabo, A., Herrup, K., Lamb, B. T., and Landreth, G. E. (2012) Ibuprofen attenuates oxidative damage through NOX2 inhibition in Alzheimer's disease. *Neurobiol. Aging* 33, 197.e121–197.e32.

(57) Yan, Q., Zhang, J., Liu, H., Babu-Khan, S., Vassar, R., Biere, A. L., Citron, M., and Landreth, G. (2003) Anti-inflammatory drug therapy alters beta-amyloid processing and deposition in an animal model of Alzheimer's disease. *J. Neurosci.* 23, 7504–7509.

(58) Czirr, E., Leuchtenberger, S., Dorner-Ciossek, C., Schneider, A., Jucker, M., Koo, E. H., Pietrzik, C. U., Baumann, K., and Weggen, S. (2007) Insensitivity to Abeta42-lowering nonsteroidal anti-inflammatory drugs and gamma-secretase inhibitors is common among aggressive presenilin-1 mutations. *J. Biol. Chem.* 282, 24504–24513.

(59) Hillmann, A., Hahn, S., Schilling, S., Hoffmann, T., Demuth, H.-U., Bulic, B., Schneider-Axmann, T., Bayer, T. A., Weggen, S., and Wirths, O. (2012) No improvement after chronic ibuprofen treatment in the 5XFAD mouse model of Alzheimer's disease. *Neurobiol. Aging* 33, 833.e39–833.e50.

(60) Kotilinek, L. A., Westerman, M. A., Wang, Q., Panizzon, K., Lim, G. P., Simonyi, A., Lesne, S., Falinska, A., Younkin, L. H., Younkin, S. G., Rowan, M., Cleary, J., Wallis, R. A., Sun, G. Y., Cole, G., Frautschy, S., Anwyl, R., and Ashe, K. H. (2008) Cyclooxygenase-2 inhibition improves amyloid-beta-mediated suppression of memory and synaptic plasticity. *Brain* 131, 651–664.

(61) Varvel, N. H., Bhaskar, K., Kounnas, M. Z., Wagner, S. L., Yang, Y., Lamb, B. T., and Herrup, K. (2009) NSAIDs prevent, but do not

reverse, neuronal cell cycle reentry in a mouse model of Alzheimer disease. *J. Clin. Invest.* 119, 3692–3702.

(62) Bombardier, C., Laine, L., Reicin, A., Shapiro, D., Burgos-Vargas, R., Davis, B., Day, R., Ferraz, M. B., Hawkey, C. J., Hochberg, M. C., Kvien, T. K., Schnitzer, T. J., and the VIGOR Study Group (2000) Comparison of upper gastrointestinal toxicity of rofecoxib and naproxen in patients with rheumatoid arthritis. *N. Engl. J. Med.* 343, 1520–1528.

(63) Egan, K. M., Lawson, J. A., Fries, S., Koller, B., Rader, D. J., Smyth, E. M., and Fitzgerald, G. A. (2004) COX-2-derived prostacyclin confers atheroprotection on female mice. *Science* 306, 1954–1957.

(64) Fitzgerald, G. A. (2004) Coxibs and cardiovascular disease. *N. Engl. J. Med.* 351, 1709–1711.

(65) Montine, T. J. T., Sidell, K. R. K., Crews, B. C. B., Markesbery, W. R. W., Marnett, L. J. L., Roberts, L. J. L., and Morrow, J. D. J. (1999) Elevated CSF prostaglandin E2 levels in patients with probable AD. *Neurology* 53, 1495–1498.

(66) Combrinck, M., Williams, J., De Berardinis, M. A., Warden, D., Puopolo, M., Smith, A. D., and Minghetti, L. (2006) Levels of CSF prostaglandin E2, cognitive decline, and survival in Alzheimer's disease. *J. Neurol., Neurosurg. Psychiatry* 77, 85–88.

(67) Breyer, R. M., Bagdasarian, C. K., Myers, S. A., and Breyer, M. D. (2001) Prostanoid receptors: subtypes and signaling. *Annu. Rev. Pharmacol. Toxicol.* 41, 661–690.

(68) Andreasson, K. (2010) Emerging roles of PGE2 receptors in models of neurological disease. *Prostaglandins Other Lipid Mediators* 91, 104–112.

(69) Johansson, J., Woodling, N., Shi, J., and Andreasson, K. (2015) Inflammatory Cyclooxygenase Activity and PGE2 Signaling in Models of Alzheimer's Disease. *Current Immunology Reviews* 11, 125–131.

(70) Carrasco, E. E., Casper, D. D., and Werner, P. P. (2007) PGE(2) receptor EP1 renders dopaminergic neurons selectively vulnerable to low-level oxidative stress and direct PGE(2) neurotoxicity. *J. Neurosci. Res.* 85, 3109–3117.

(71) Zhen, G., Kim, Y. T., Li, R.-C., Yocum, J., Kapoor, N., Langer, J., Dobrowolski, P., Maruyama, T., Narumiya, S., and Doré, S. (2012) PGE2 EP1 receptor exacerbated neurotoxicity in a mouse model of cerebral ischemia and Alzheimer's disease. *Neurobiol. Aging* 33, 2215–2219.

(72) Liang, X., Wang, Q., Hand, T., Wu, L., Breyer, R. M., Montine, T. J., and Andreasson, K. (2005) Deletion of the prostaglandin E2 EP2 receptor reduces oxidative damage and amyloid burden in a model of Alzheimer's disease. *J. Neurosci.* 25, 10180–10187.

(73) Tamagno, E. E., Bardini, P. P., Obbili, A. A., Vitali, A. A., Borghi, R. R., Zaccheo, D. D., Pronzato, M. A. M., Danni, O. O., Smith, M. A. M., Perry, G. G., and Tabaton, M. M. (2002) Oxidative stress increases expression and activity of BACE in NT2 neurons. *Neurobiol. Dis.* 10, 279–288.

(74) Montine, T. J., Milatovic, D., Gupta, R. C., Valyi-Nagy, T., Morrow, J. D., and Breyer, R. M. (2002) Neuronal oxidative damage from activated innate immunity is EP2 receptor-dependent. *J. Neurochem.* 83, 463–470.

(75) Shie, F.-S., Montine, K. S., Breyer, R. M., and Montine, T. J. (2005) Microglial EP2 is critical to neurotoxicity from activated cerebral innate immunity. *Glia* 52, 70–77.

(76) Shie, F.-S., Breyer, R. M., and Montine, T. J. (2005) Microglia lacking E Prostanoid Receptor subtype 2 have enhanced Abeta phagocytosis yet lack Abeta-activated neurotoxicity. *Am. J. Pathol.* 166, 1163–1172.

(77) Keene, C. D., Chang, R. C., Lopez-Yglesias, A. H., Shalloway, B. R., Sokal, I., Li, X., Reed, P. J., Keene, L. M., Montine, K. S., Breyer, R. M., Rockhill, J. K., and Montine, T. J. (2010) Suppressed Accumulation of Cerebral Amyloid {beta} Peptides in Aged Transgenic Alzheimer's Disease Mice by Transplantation with Wild-Type or Prostaglandin E2 Receptor Subtype 2-Null Bone Marrow. *Am. J. Pathol.* 177, 346.

(78) Li, X., Melief, E., Postupna, N., Montine, K. S., Keene, C. D., and Montine, T. J. (2015) Prostaglandin E2 receptor subtype 2

regulation of scavenger receptor CD36 modulates microglial A $\beta$ 42 phagocytosis. *Am. J. Pathol.* 185, 230–239.

(79) Johansson, J. U., Pradhan, S., Lokteva, L. A., Woodling, N. S., Ko, N., Brown, H. D., Wang, Q., Loh, C., Cekanaviciute, E., Buckwalter, M., Manning-Boğ, A. B., and Andreasson, K. I. (2013) Suppression of inflammation with conditional deletion of the prostaglandin E2 EP2 receptor in macrophages and brain microglia. *J. Neurosci.* 33, 16016–16032.

(80) Johansson, J. U., Woodling, N. S., Wang, Q., Panchal, M., Liang, X., Trueba-Saiz, A., Brown, H. D., Mhatre, S. D., Loui, T., and Andreasson, K. I. (2015) Prostaglandin signaling suppresses beneficial microglial function in Alzheimer's disease models. *J. Clin. Invest.* 125, 350–364.

(81) Shi, J., Wang, Q., Johansson, J. U., Liang, X., Woodling, N. S., Priyam, P., Loui, T. M., Merchant, M., Breyer, R. M., Montine, T. J., and Andreasson, K. (2012) Inflammatory prostaglandin E2 signaling in a mouse model of Alzheimer disease. *Ann. Neurol.* 72, 788–798.

(82) Liang, X., Lin, L., Woodling, N. S., Wang, Q., Anacker, C., Pan, T., Merchant, M., and Andreasson, K. (2011) Signaling via the prostaglandin E<sub>2</sub> receptor EP4 exerts neuronal and vascular protection in a mouse model of cerebral ischemia. *J. Clin. Invest.* 121, 4362–4371.

(83) Minami, M., Shimizu, K., Okamoto, Y., Folco, E., Ilasaca, M.-L., Feinberg, M. W., Aikawa, M., and Libby, P. (2008) Prostaglandin E receptor type 4-associated protein interacts directly with NF- $\kappa$ B1 and attenuates macrophage activation. *J. Biol. Chem.* 283, 9692–9703.

(84) Shi, J., Johansson, J., Woodling, N. S., Wang, Q., Montine, T. J., and Andreasson, K. (2010) The prostaglandin E2 E-prostanoid 4 receptor exerts anti-inflammatory effects in brain innate immunity. *J. Immunol.* 184, 7207–7218.

(85) Kabashima, K. K., Saji, T. T., Murata, T. T., Nagamachi, M. M., Matsuoka, T. T., Segi, E. E., Tsuboi, K. K., Sugimoto, Y. Y., Kobayashi, T. T., Miyachi, Y. Y., Ichikawa, A. A., and Narumiya, S. S. (2002) The prostaglandin receptor EP4 suppresses colitis, mucosal damage and CD4 cell activation in the gut. *J. Clin. Invest.* 109, 883–893.

(86) Yamagata, K., Andreasson, K. I., Kaufmann, W. E., Barnes, C. A., and Worley, P. F. (1993) Expression of a mitogen-inducible cyclooxygenase in brain neurons: regulation by synaptic activity and glucocorticoids. *Neuron* 11, 371–386.

(87) Melnikova, T., Savonenko, A., Wang, Q., Liang, X., Hand, T., Wu, L., Kaufmann, W. E., Vehmas, A., and Andreasson, K. I. (2006) Cyclooxygenase-2 activity promotes cognitive deficits but not increased amyloid burden in a model of Alzheimer's disease in a sex-dimorphic pattern. *Neuroscience* 141, 1149–1162.

(88) Lim, G. (2001) Ibuprofen effects on Alzheimer pathology and open field activity in APPsw transgenic mice. *Neurobiol. Aging* 22, 983–991.

(89) Jantzen, P. T., Connor, K. E., DiCarlo, G., Wenk, G. L., Wallace, J. L., Rojiani, A. M., Coppola, D., Morgan, D., and Gordon, M. N. (2002) Microglial activation and beta -amyloid deposit reduction caused by a nitric oxide-releasing nonsteroidal anti-inflammatory drug in amyloid precursor protein plus presenilin-1 transgenic mice. *J. Neurosci.* 22, 2246–2254.

(90) Eriksen, J. L., Sagi, S. A., Smith, T. E., Weggen, S., Das, P., McLendon, D. C., Ozols, V. V., Jessing, K. W., Zavitz, K. H., Koo, E. H., and Golde, T. E. (2003) NSAIDs and enantiomers of flurbiprofen target gamma-secretase and lower Abeta 42 in vivo. *J. Clin. Invest.* 112, 440–449.

(91) Quinn, J., Montine, T., Morrow, J., Woodward, W. R., Kulhanek, D., and Eckenstein, F. (2003) Inflammation and cerebral amyloidosis are disconnected in an animal model of Alzheimer's disease. *J. Neuroimmunol.* 137, 32–41.

(92) Sung, S., Yang, H., Uryu, K., Lee, E. B., Zhao, L., Shineman, D., Trojanowski, J. Q., Lee, V. M.-Y., and Pratico, D. (2004) Modulation of Nuclear Factor- $\kappa$ B Activity by Indomethacin Influences A $\beta$  Levels but Not A $\beta$  Precursor Protein Metabolism in a Model of Alzheimer's Disease. *Am. J. Pathol.* 165, 2197–2206.

(93) Lanz, T. A., Fici, G. J., and Merchant, K. M. (2005) Lack of specific amyloid-beta(1–42) suppression by nonsteroidal anti-

inflammatory drugs in young, plaque-free Tg2576 mice and in guinea pig neuronal cultures. *J. Pharmacol. Exp. Ther.* 312, 399–406.

(94) van Groen, T., and Kadish, I. (2005) Transgenic AD model mice, effects of potential anti-AD treatments on inflammation and pathology. *Brain Res. Rev.* 48, 370–378.

(95) Kukar, T., Prescott, S., Eriksen, J. L., Holloway, V., Murphy, M. P., Koo, E. H., Golde, T. E., and Nicolle, M. M. (2007) Chronic administration of R-flurbiprofen attenuates learning impairments in transgenic amyloid precursor protein mice. *BMC Neurosci.* 8, 54.

(96) McKee, A. C., Carreras, I., Hossain, L., Ryu, H., Klein, W. L., Oddo, S., LaFerla, F. M., Jenkins, B. G., Kowall, N. W., and Dedeoglu, A. (2008) Ibuprofen reduces Abeta, hyperphosphorylated tau and memory deficits in Alzheimer mice. *Brain Res.* 1207, 225–236.

(97) Carreras, I., McKee, A. C., Choi, J.-K., Aytan, N., Kowall, N. W., Jenkins, B. G., and Dedeoglu, A. (2013) R-flurbiprofen improves tau, but not A $\beta$  pathology in a triple transgenic model of Alzheimer's disease. *Brain Res.* 1541, 115–127.

(98) Choi, S.-H., Aid, S., Caracciolo, L., Minami, S. S., Niikura, T., Matsuoka, Y., Turner, R. S., Mattson, M. P., and Bosetti, F. (2013) Cyclooxygenase-1 inhibition reduces amyloid pathology and improves memory deficits in a mouse model of Alzheimer's disease. *J. Neurochem.* 124, 59–68.

(99) Subaiea, G. M., Adwan, L. I., Ahmed, A. H., Stevens, K. E., and Zawia, N. H. (2013) Short-term treatment with tolfenamic acid improves cognitive functions in Alzheimer's disease mice. *Neurobiol. Aging* 34, 2421–2430.

(100) Choi, J.-K., Carreras, I., Aytan, N., Jenkins-Sahlin, E., Dedeoglu, A., and Jenkins, B. G. (2014) The effects of aging, housing and ibuprofen treatment on brain neurochemistry in a triple transgene Alzheimer's disease mouse model using magnetic resonance spectroscopy and imaging. *Brain Res.* 1590, 85–96.